

# Review of the Pathophysiology of Autism and Possible Benefits of Hyperbaric Oxygen Therapy (HBOT)

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## HBOT and Autism Overview

At first glance, the use of hyperbaric oxygen therapy (HBOT) in autism appears out of the ordinary. That is what I first thought when I heard about the use of HBOT in autism almost 2 years ago. At the time, no studies existed on the use of HBOT in individuals with autism (there was one published case report [1]). In fact, many people who were proponents for this therapy could not give a theoretical reason why it should/could work. We began using HBOT in children with autism with a great deal of skepticism. After seeing improvements in some of these children (including our own), we decided that further study was needed. Just over 2 years later, we have finished our third study on the use of HBOT in autism. Many of the underlying pathophysiological findings in autism might be improved with HBOT and these have been reviewed in another publication [2]. HBOT in children is generally regarded as safe [3].

## Cerebral Hypoperfusion and Hypoxia in Autism

To understand how or why HBOT works in children with autism, we need to review some basic, but newly described, fundamental problems found in many individuals with autism. There are now numerous studies in the medical literature [4-11] demonstrating cerebral hypoperfusion (decreased blood flow to the brain) in as many as 86% of individuals with autism [4]. In one study, this hypoperfusion typically worsened as the age of the child increased, and become “quite profound” in older children compared to younger [5]. Furthermore, this diminished blood flow typically correlates with many core autistic symptoms (see *Table 1*).

When a typical person has to focus on a task or generate speech (in other words, when the brain has to do more work), there is an increase in blood flow to the brain, supplying more blood, oxygen, and glucose (fuel) [12]. However, several studies have now demonstrated that not only do some

children with autism have diminished blood flow at baseline, they also do not get an increase in blood flow when brain cells have to do more work, such as when the children have to pay attention to a task or generate a sentence [13-15]. In fact, sometimes the cerebral blood flow actually goes *down*, and this appears to be mediated, in part, by inappropriate vasoconstriction (narrowing of blood vessels) instead of vasodilatation [15]. The interesting thing about these studies demonstrating cerebral hypoperfusion in autism is that none of these studies has stopped to examine *why* the diminished blood flow exists in the first place. This cerebral hypoperfusion appears to lead to cerebral hypoxia (impaired oxygen delivery) to the brain in some individuals with autism. In fact, several studies have demonstrated a reduction in Bcl-2 and an increase in p53 in the brain of some children with autism [16, 17]. Elevated p53 is caused by hypoxia [18] and an increase in Bcl-2 normally protects from cell death provoked by hypoxia; a reduction is associated with increased damage caused by hypoxia [19].

**Table 1: Selected Areas of Cerebral Hypoperfusion in Autism and Clinical Correlations**

<i>Area of Cerebral Hypoperfusion</i>	<i>Clinical Correlation</i>
Thalamus	Repetitive, self-stimulatory, and unusual behaviors [6]
Temporal lobes	Desire for sameness and social/communication impairments [7]
Temporal lobes and amygdala	Impairments in processing facial expressions and emotions [8]
Fusiform gyrus	Difficulty recognizing familiar faces [9]
Wernicke’s and Brodmann’s areas	Decreased language development and auditory processing problems [5; 10]
Temporal and frontal lobes	Decreased IQ [11]

## **Cerebral Hypoperfusion and Neuroinflammation in Autism**

The cause of cerebral hypoperfusion in individuals with autism is unknown but might be due to inflammation. Evidence published out of Johns Hopkins in 2005 demonstrates that, upon autopsy, some individuals with autism have evidence of inflammation in the brain (neuroinflammation) [20]. Also described was inflammation around blood vessels, which is consistent with vasculitis (inflamed blood vessels), and could cause the vessel wall to become stiff and inflexible. Vasculitis decreases the ability of the blood vessel to dilate and can lead to diminished blood flow. There have been several other studies in the literature confirming the presence of inflammation in the brain of some individuals with autism [21-23]. Children with autism also produce more serum autoantibodies to the brain [24], including IgG and IgM autoantibodies to brain endothelial cells and nuclei when compared to typical children [25]. Elevated serum autoantibodies to many neuron-specific antigens and cross-reactive peptides have been found in children with autism [26], including antibodies directed against cerebellar Purkinje cells [27] and neural proteins such as myelin basic protein [26, 28]. Furthermore, in one study, 49% of children with autism created serum antibodies against the caudate nucleus and 18% produced serum antibodies to the cerebral cortex [29].

Inflammation generally is associated with edema (increased swelling), can increase the space between cells [30], and might increase the amount of fluid present inside cells. Two fMRI (functional Magnetic Resonance Imaging) studies published in 2006 demonstrated that individuals with autism had more fluid inside brain cells when compared to typical children [31-32]. Furthermore, functional connectivity (the ability of one brain cell to communicate to another cell) is diminished in some children with autism when compared to typical children [33]. It is possible that inflammation present in the brain of some individuals with autism leads to diminished blood flow, impaired functional connectivity, and increased fluid inside brain cells as described in these studies.

Furthermore, elevated urinary levels of 8-isoprostane-F2 $\alpha$  have recently been described in some individuals with autism [34]. In some studies, this isoprostane elevation has been shown to cause *in vivo* vasoconstriction and increase the aggrega-

tion (stickiness) of platelets. A more recent study also demonstrated increased urinary levels of isoprostane F2 $\alpha$ -VI (a marker of lipid peroxidation, or oxidative stress), 2,3-dinor-thromboxane B2 (which reflects platelet activation), and 6-keto-prostaglandin F1 $\alpha$  (a marker of endothelium activation) [35]. Therefore, the inflammation surrounding blood vessels, and the increase in the inflammatory substances leading to vasoconstriction, and increased activation of platelets and endothelium might cause the diminished cerebral blood flow found in many individuals with autism.

Treatment of this inflammation might help restore normal blood flow. In fact, many inflammatory conditions such as lupus, Kawasaki disease, Behçet's disease, encephalitis, and Sjögren's syndrome are characterized by cerebral hypoperfusion [36-42], and treatment with anti-inflammatory medication can restore normal cerebral blood flow in some of these conditions [43, 44]. In addition, review of the literature demonstrates that the use of anti-inflammatory treatments appears to improve symptoms in some children with autism [45]. In fact, treatment with corticosteroids in one child who developed an autoimmune lymphoproliferative syndrome and subsequent autism led to objective improvements in speech and developmental milestones [46]. In another child with autism, whose behavior and language regressed at 22 months of age, treatment with corticosteroids ameliorated abnormal behaviors such as hyperactivity, tantrums, impaired social interaction, echolalia, and stereotypies [47].

## **Gastrointestinal Inflammation in Autism**

Also described in dozens of studies is the presence of inflammation in the gastrointestinal tract of some children with autism. This has been termed autistic enterocolitis or chronic ileocolonic lymphoid nodular hyperplasia (LNH). This condition is characterized by mucosal inflammation of the colon, stomach, and small intestine [48-50]. As many as 90% of children with autism with gastrointestinal symptoms (diarrhea, constipation, etc...) have evidence of ileal LNH, with 68% having moderate to severe ileal LNH [48]. Several studies have shown that some children with autism have evidence of inflammatory cells such as lymphocytes [51, 52] and eosinophils [53] inside the gastrointestinal mucosa, sometimes mimicking an autoimmune lesion [51]. Inflammatory markers in the blood are also elevated, including TNF- $\alpha$ , Interferon- $\gamma$  (IFN- $\gamma$ ), and IL-6, and anti-inflammatory markers such as IL-10 are decreased

[52, 54]. Children with autism typically make significantly more serum antibodies against gliadin and casein peptides resulting in autoimmune reactions [55]. More than 25% of individuals with autism make serum IgG, IgM, and IgA antibodies against gliadin which can then cross-react with cerebellar peptides [23].

### **HBOT, Cerebral Hypoperfusion, and Inflammation**

Since the cerebral hypoperfusion in autism is likely secondary to inflammation, HBOT might be especially helpful because it possesses potent anti-inflammatory tissue effects [56], with equivalence to diclofenac (an anti-inflammatory medication) 20 mg/kg noted in one animal study [57]. HBOT has been used in cases of vasculitis with good results [58], and with some success in disorders characterized by cerebral hypoperfusion including fetal alcohol syndrome [59], cerebral palsy [60, 61], chronic brain injury [62], closed head injury [63], and stroke [64]. HBOT also attenuated the production of proinflammatory cytokines including TNF- $\alpha$  [65], IL-1 [65], IL-1 $\beta$  [66], and IL-6 [65],

and increased the production of anti-inflammatory IL-10 [67]. HBOT reduced neuroinflammation in a rat model after traumatic brain injury [68], and diminished both inflammation and pain in an animal model of inflammatory pain [69]. HBOT has been used in humans to achieve remission of Crohn’s disease [70-74] and ulcerative colitis [75, 76] not responding to conventional medications, including corticosteroids and immunosuppressive drugs. Interestingly, in some studies, the decrease in inflammation with HBOT appeared to be caused by the increased pressure, not necessarily by the increased oxygen tension. In one animal study, hyperbaric pressure without additional oxygen was shown to decrease TNF- $\alpha$  levels [77]. In a human study, HBOT at 2 atmosphere (atm) and 100% oxygen, and hyperbaric pressure at 2 atm and 10.5% oxygen (thus supplying 21% oxygen, equal to room air oxygen levels) both showed anti-inflammatory activity by inhibiting IFN- $\gamma$  release, whereas 100% oxygen at room air pressure (1 atm) actually increased IFN- $\gamma$  release [78]. For these reasons, HBOT might help ameliorate the inflammation found in some individuals with autism (see *Table 2*).

**Table 2: Effects of HBOT on Inflammatory Markers and Inflammation in Autism**

<i>Marker</i>	<i>Classification</i>	<i>Autism Finding</i>	<i>HBOT Effect</i>
TNF- $\alpha$	Inflammatory	↑ [52; 54]	↓ [65; 66; 77 <sup>1</sup> ]
IL-1 $\beta$	Inflammatory	↑ [54]	↓ [66]
IL-6	Inflammatory	↑ [20; 54]	↓ [65]
IL-10	Anti-inflammatory	↓ [52]	↑ [67]
IFN- $\gamma$	Inflammatory	↑ [52]	↓ [78 <sup>2</sup> ]
Neuroinflammation		↑ [20-22]	↓ [65]
GI inflammation		↑ [48-50]	↓ [70; 75]

<sup>1</sup> Hyperbaric pressure without additional oxygen decreased TNF- $\alpha$ .

<sup>2</sup> Hyperbaric pressure without additional oxygen also decreased IFN- $\gamma$ .

### **Clinical Studies on HBOT in Autism**

In one case report, Heuser et al. treated a four year old child with autism using hyperbaric therapy at 1.3 atm and 24% oxygen and reported “striking improvement in behavior including memory and cognitive functions” after only ten sessions. The child also had marked improvement of cerebral hypoperfusion as measured by pre-hyperbaric and post-hyperbaric Single Photon Emission Computed Tomography (SPECT) scans [1]. Our previous case series suggested that hyperbaric therapy at 1.3 atm and 28% oxygen led to clinical improvements in some children with

autism as measured by the Autism Treatment Evaluation Checklist (ATEC), Childhood Autism Rating Scale (CARS), and Social Responsiveness Scale (SRS) scales [79]. This low pressure HBOT was well tolerated by all 6 children with no adverse effects noted.

Recently published is a prospective open-label study on 18 children with autism who underwent 40 hyperbaric sessions of 45 minutes duration each at either 1.5 atm and 100% oxygen (6 children), or 1.3 atm and 24% oxygen (12 children) [80]. Results were calculated before and after the 40 treatments using parent-rated Aberrant Behavior Checklist-Community

(ABC-C), SRS, and ATEC. Fasting blood was drawn before and after the 40 treatments for C-reactive protein (CRP) and markers of oxidative stress.

For the 1.5 atm group, parents reported significant improvements in irritability, lethargy, hyperactivity, motivation, and sensory and cognitive awareness. For the 1.3 atm group, parents reported significant improvements in motivation, mannerisms, physical health, sensory and cognitive awareness, speech, and communication. Mean CRP improved in both groups, especially in a subgroup of children with very elevated initial CRP. There was no statistically significant change in mean plasma oxidized glutathione levels in either group after 40 treatments. Oxidized glutathione is normally expelled out of cells when oxidative stress inside the cell is high [81]. Therefore, since oxidized glutathione did not appreciably increase in the plasma, oxidative stress did not significantly worsen at either pressure in this study. Comparisons between the 2 groups in this study in clinical outcomes must be done with caution because of the small number of participants involved. Finally, HBOT at both 1.5 atm/100% oxygen and 1.3 atm/24% oxygen were both used safely in children with autism.

## References

- [1] Heuser G, Heuser SA, Rodeland D, Aguilera O, Uszler M. Treatment of neurologically impaired adults and children with "mild" hyperbaric oxygenation (1.3 ATM and 24% oxygen). In *Hyperbaric oxygenation for cerebral palsy and the brain-injured child*. Edited by Joiner JT. Flagstaff Arizona: Best Publications; 2002:109-15.
- [2] Rossignol DA. Hyperbaric oxygen therapy might improve certain pathophysiological findings in autism. *Med Hypotheses* 2006, in press.
- [3] Ashamalla HL, Thom SR, Goldwein JW. Hyperbaric oxygen therapy for the treatment of radiation-induced sequelae in children: the University of Pennsylvania experience. *Cancer* 1996;77(11):2407-12.
- [4] Zilbovicius M, Boddart N, Belin P, et al. Temporal lobe dysfunction in childhood autism: a PET study. *Am J Psychiatry* 2000;157(12):1988-93.
- [5] Wilcox J, Tsuang MT, Ledger E, Algeo J, Schnurr T. Brain perfusion in autism varies with age. *Neuropsychobiology* 2002;46(1):13-6.
- [6] Starkstein SE, Vazquez S, Vrancic D, et al. SPECT findings in mentally retarded autistic individuals. *J Neuropsychiatry Clin Neurosci* 2000;12(3):370-5.
- [7] Ohnishi T, Matsuda H, Hashimoto T, et al. Abnormal regional cerebral blood flow in childhood autism. *Brain* 2000;123(Pt9):1838-44.
- [8] Critchley HD, Daly EM, Bullmore ET, et al. The functional neuroanatomy of social behaviour: changes in cerebral blood flow when people with autistic disorder process facial expressions. *Brain* 2000;123(Pt11):2203-12.
- [9] Pierce K, Haist F, Sedaghat F, Courchesne E. The brain response to personally familiar faces in autism: findings of fusiform activity and beyond. *Brain* 2004;127(Pt12):2703-16.
- [10] Boddart N, Zilbovicius M. Functional neuroimaging and childhood autism. *Pediatr Radiol* 2002;32(1):1-7.
- [11] Hashimoto T, Sasaki M, Fukumizu M, Hanaoka S, Sugai K, Matsuda H. Single-photon emission computed tomography of the brain in autism: effect of the developmental level. *Pediatr Neurol* 2000;23(5):416-20.
- [12] Fox PT, Raichle ME. Focal physiological uncoupling of cerebral blood flow and oxidative metabolism during somatosensory stimulation in human subjects. *Proc Natl Acad Sci USA* 1986;83(4):1140-4.
- [13] Allen G, Courchesne E. Differential effects of developmental cerebellar abnormality on cognitive and motor functions in the cerebellum: an fMRI study of autism. *Am J Psychiatry* 2003;160(2):262-73.
- [14] Muller RA, Behen ME, Rothermel RD, et al. Brain mapping of language and auditory perception in high-functioning autistic adults: a PET study. *J Autism Dev Disord* 1999;29(1):19-31.
- [15] Bruneau N, Dourneau MC, Garreau B, Pourcelot L, Lelord G. Blood flow response to auditory stimulations in normal, mentally retarded, and autistic children: a preliminary transcranial Doppler ultrasonographic study of the middle cerebral arteries. *Biol Psych* 1992;32(8):691-9.
- [16] Fatemi, S.H., A.R. Halt, 2001. Altered levels of Bcl2 and p53 proteins in parietal cortex reflect deranged apoptotic regulation in autism. *Synapse*, 42(4):281-4.
- [17] Araghi-Niknam, M., S.H Fatemi, 2003. Levels of Bcl-2 and P53 are altered in superior frontal and cerebellar cortices of autistic subjects. *Cell Mol. Neurobiol.*, 23(6):945-52.
- [18] Graeber, T.G., J.F. Peterson, M. Tsai, K. Monica, A.J, Fornace Jr, A.J Giaccia, 1994. Hypoxia induces accumulation of p53 protein, but activation of a G1-phase checkpoint by low-oxygen conditions is independent of p53 status. *Mol. Cell Biol.*, 14(9):6264-77.
- [19] Shimizu, S., Y. Eguchi, W. Kamiike, Y. Itoh, J. Hasegawa, K. Yamabe, Y. Otsuki, H. Matsuda, Y. Tsujimoto, 1996. Induction of apoptosis as well as necrosis by hypoxia and predominant prevention of apoptosis by Bcl-2 and Bcl-XL. *Cancer Res.*, 56(9):2161-6.
- [20] Vargas DL, Nascimbene C, Krishnan C, Zimmerman AW, Pardo CA. Neuroglial activation and neuroinflammation in the brain of patients with autism. *Ann Neurol* 2005;57(1):67-81.
- [21] Pardo CA, Vargas DL, Zimmerman AW. Immunity, neuroglia and neuroinflammation in autism. *Int Rev Psychiatry* 2005;17(6):485-95.
- [22] Laurence JA, Fatemi SH. Glial fibrillary acidic protein is elevated in superior frontal, parietal and cerebellar cortices of autistic subjects. *Cerebellum* 2005;4(3):206-10.

- [23] Weizman A, Wiezman R, Szekely GA, Wijzenbeek H, Livni E. Abnormal immune response to brain tissue antigen in the syndrome of autism. *Am J Psychiatry* 1982;139(11):1462-5.
- [24] Singer HS, Morris CM, Williams PN, Yoon DY, Hong JJ, Zimmerman AW. Antibrain antibodies in children with autism and their unaffected siblings. *J Neuroimmunol* 2006;178(1-2):149-55.
- [25] Connolly AM, Chez MG, Pestronk A, Arnold ST, Mehta S, Deuel RK. Serum autoantibodies to brain in Landau-Kleffner variant, autism, and other neurologic disorders. *J Pediatr* 1999;134(5):607-13.
- [26] Vojdani A, Campbell AW, Anyanwu E, Kashanian A, Bock K, Vojdani E. Antibodies to neuron-specific antigens in children with autism: possible cross-reaction with encephalitogenic proteins from milk, Chlamydia pneumoniae and Streptococcus group A. *J Neuroimmunol* 2002;129(1-2):168-77.
- [27] Vojdani A, O'Bryan T, Green JA, et al. Immune response to dietary proteins, gliadin and cerebellar peptides in children with autism. *Nutr Neurosci* 2004;7(3):151-61.
- [28] Singh VK, Warren RP, Odell JD, Warren WL, Cole P. Antibodies to myelin basic protein in children with autistic behavior. *Brain Behav Immun* 1993;7(1):97-103.
- [29] Singh VK, Rivas WH. Prevalence of serum antibodies to caudate nucleus in autistic children. *Neurosci Lett* 2004;355(1-2):53-6.
- [30] Lu G, Qian X, Berezin I, Telford GL, Huizinga JD, Sarna SK. Inflammation modulates in vitro colonic myoelectric and contractile activity and interstitial cells of Cajal. *Am J Physiol* 1997;273(6 Pt 1):G1233-45.
- [31] Hendry J, DeVito T, Gelman N, Densmore M, Rajakumar N, Pavlosky W, Williamson PC, Thompson PM, Drost DJ, Nicolson R. White matter abnormalities in autism detected through transverse relaxation time imaging. *Neuroimage* 2006;29(4):1049-57.
- [32] Petropoulos H, Friedman SD, Shaw DW, Artru AA, Dawson G, Dager SR. Gray matter abnormalities in autism spectrum disorder revealed by T2 relaxation. *Neurology* 2006;67(4):632-6.
- [33] Just MA, Cherkassky VL, Keller TA, Minshew NJ. Cortical activation and synchronization during sentence comprehension in high-functioning autism: evidence of underconnectivity. *Brain* 2004;127(Pt 8):1811-21.
- [34] Ming X, Stein TP, Brimacombe M, Johnson WG, Lambert GH, Wagner GC. Increased excretion of a lipid peroxidation biomarker in autism. *Prostaglandins Leukot Essent Fatty Acids* 2005;73(5):379-84.
- [35] Yao Y, Walsh WJ, McGinnis WR, Pratico D. Altered vascular phenotype in autism: correlation with oxidative stress. *Arch Neurol* 2006;63(8):1161-4.
- [36] Ichiyama T, Nishikawa M, Hayashi T, Koga M, Tashiro N, Furukawa S. Cerebral hypoperfusion during acute Kawasaki Disease. *Stroke* 1998;29(7):1320-1.
- [37] Huang WS, Chiu PY, Tsai CH, Kao A, Lee CC. Objective evidence of abnormal regional cerebral blood flow in patients with systemic lupus erythematosus on Tc-99m ECD brain SPECT. *Rheumatol Int* 2002;22(5):178-181.
- [38] Postiglione A, De Chiara S, Soricelli A, et al. Alterations of cerebral blood flow and antiphospholipid antibodies in patients with systemic lupus erythematosus. *Int J Clin Lab Res* 1998;28(1):34-8.
- [39] Lass P, Krajka-Lauer J, Homziuk M, et al. Cerebral blood flow in Sjögren's syndrome using 99Tcm-HMPAO brain SPET. *Nucl Med Commun* 2000;21(1):31-5.
- [40] Caca I, Nazaroglu H, Unlu K, Cakmak SS, Ari S, Sakalar YB. Color Doppler imaging of ocular hemodynamic changes in Behçet's disease. *Jpn J Ophthalmol* 2004;48(2):101-5.
- [41] Wakamoto H, Ohta M, Nakano N, Kunisue K. SPECT in focal enterovirus encephalitis: evidence for local cerebral vasculitis. *Pediatr Neurol* 2000;23(5):429-31.
- [42] Nishikawa M, Matsubara T, Yoshitomi T, Ichiyama T, Hayashi T, Furukawa S. Abnormalities of brain perfusion in echovirus type 30 meningitis. *J Neurol Sci* 2000;179(S1-2):122-6.
- [43] Mathieu A, Sanna G, Marni A, et al. Sustained normalization of cerebral blood-flow after iloprost therapy in a patient with neuropsychiatric systemic lupus erythematosus. *Lupus* 2002;11(1):52-6.
- [44] Liu FY, Huang WS, Kao CH, Yen RF, Wang JJ, Ho ST. Usefulness of Tc-99m ECD brain SPECT to evaluate the effects of methylprednisolone pulse therapy in lupus erythematosus with brain involvement: a preliminary report. *Rheumatol Int* 2003;23(4):182-5.
- [45] Wakefield AJ, Puleston JM, Montgomery SM, Anthony A, O'Leary JJ, Murch SH. Review article: the concept of entero-colonic encephalopathy, autism and opioid receptor ligands. *Aliment Pharmacol Ther* 2002;16(4):663-74.
- [46] Shenoy S, Arnold S, Chatila T. Response to steroid therapy in autism secondary to autoimmune lymphoproliferative syndrome. *J Pediatr* 2000;136(5):682-7.
- [47] Stefanatos GA, Grover W, Geller E. Case study: corticosteroid treatment of language regression in pervasive developmental disorder. *J Am Acad Child Adolesc Psychiatry* 1995;34(8):1107-11.
- [48] Wakefield AJ, Ashwood P, Limb K, Anthony A. The significance of ileo-colonic lymphoid nodular hyperplasia in children with autistic spectrum disorder. *Eur J Gastroenterol Hepatol* 2005;17(8):827-36.
- [49] Uhlmann V, Martin CM, Sheils O, et al. Potential viral pathogenic mechanism for new variant inflammatory bowel disease. *J Clin Pathol: Mol Pathol* 2002;55(2):84-90.
- [50] Furlano RI, Anthony A, Day R, et al. Colonic CD8 and gamma delta T-cell infiltration with epithelial damage in children with autism. *J Pediatr* 2001;138(3):366-72.
- [51] Torrente F, Ashwood P, Day R, et al. Small intestinal enteropathy with epithelial IgG and complement deposition in children with regressive autism. *Mol Psychiatry* 2002;7(4):375-82.
- [52] Ashwood P, Anthony A, Torrente F, Wakefield AJ. Spontaneous mucosal lymphocyte cytokine profiles in children with autism and gastrointestinal symptoms: mucosal immune activation and reduced counter regulatory interleukin-10. *J Clin Immunol* 2004;24(6):664-73.
- [53] Ashwood P, Anthony A, Pellicer AA, Torrente F, Walker-Smith JA, Wakefield AJ. Intestinal lymphocyte populations in children with regressive autism: evidence for extensive mucosal immunopathology. *J Clin Immunol* 2003;23(6):504-17.
- [54] Jyonouchi H, Sun S, Le H. Proinflammatory and regulatory cytokine production associated with innate and adaptive im-

- immune responses in children with autism spectrum disorders and developmental regression. *J Neuroimmunol* 2001;120(1-2):170-9.
- [55] Vojdani A, Pangborn JB, Vojdani E, Cooper EL. Infections, toxic chemicals and dietary peptides binding to lymphocyte receptors and tissue enzymes are major instigators of autoimmunity in autism. *Int J Immunopathol Pharmacol* 2003;16(3):189-99.
- [56] Al-Waili NS, Butler GJ. Effects of hyperbaric oxygen on inflammatory response to wound and trauma: possible mechanism of action. *Scient World Journal* 2006;6:425-41
- [57] Sumen G, Cimsit M, Eroglu L. Hyperbaric oxygen treatment reduces carrageenan-induced acute inflammation in rats. *Eur J Pharmacol* 2001;431(2):265-8.
- [58] Efrati S, Bergan J, Fishlev G, Tishler M, Golik A, Gall N. Hyperbaric oxygen therapy for nonhealing vasculitic ulcers. *Clinical Dermatology* 2007;32(1):12-7.
- [59] Stoller KP. Quantification of neurocognitive changes before, during, and after hyperbaric oxygen therapy in a case of fetal alcohol syndrome. *Pediatrics* 2005;116(4):e586-91.
- [60] Montgomery D, Goldberg J, Amar M, et al. Effects of hyperbaric oxygen therapy on children with spastic diplegic cerebral palsy: a pilot project. *Undersea Hyperb Med* 1999;26(4):235-42.
- [61] Collet JP, Vanasse M, Marois P, et al. Hyperbaric oxygen for children with cerebral palsy: a randomised multicentre trial. *Lancet* 2001;357(9256):582-6.
- [62] Golden ZL, Neubauer R, Golden CJ, Greene L, Marsh J, Mleko A. Improvement in cerebral metabolism in chronic brain injury after hyperbaric oxygen therapy. *Int J Neurosci* 2002;112(2):119-31.
- [63] Rockswold GL, Ford SE, Anderson DC, Bergman TA, Sherman RE. Results of a prospective randomized trial for the treatment of severely brain-injured patients with hyperbaric oxygen. *J Neurosurg* 1992;76(6):929-34.
- [64] Nighoghossian N, Trouillas P, Adeleine P, Salord F. Hyperbaric oxygen in the treatment of acute ischemic stroke. *Stroke* 1995;26:1369-72.
- [65] Weisz G, Lavy A, Adir Y, et al. Modification of in vivo and in vitro TNF-alpha, IL-1, and IL-6 secretion by circulating monocytes during hyperbaric oxygen treatment in patients with perianal Crohn's disease. *J Clin Immunol* 1997;17(2):154-9.
- [66] Benson RM, Minter LM, Osborne BA, Granowitz EV. Hyperbaric oxygen inhibits stimulus-induced proinflammatory cytokine synthesis by human blood-derived monocyte-macrophages. *Clin Exp Immunol* 2003;134(1):57-62.
- [67] Buras JA, Holt D, Orlow D, Belikoff B, Pavlides S, Reenstra WR. Hyperbaric oxygen protects from sepsis mortality via an interleukin-10-dependent mechanism. *Crit Care Med* 2006;34(10):2624-9.
- [68] Vlodavsky E, Palzur E, Soustiel JF. Hyperbaric oxygen therapy reduces neuroinflammation and expression of matrix metalloproteinase-9 in the rat model of traumatic brain injury. *Neuropathol Appl Neurobiol* 2006;32(1):40-50.
- [69] Wilson HD, Wilson JR, Fuchs PN. Hyperbaric oxygen treatment decreases inflammation and mechanical hypersensitivity in an animal model of inflammatory pain. *Brain Res* 2006;1098(1):126-8.
- [70] Takeshima F, Makiyama K, Doi T. Hyperbaric oxygen as adjunct therapy for Crohn's intractable enteric ulcer. *Am J Gastroenterol* 1999;94(11):3374-5.
- [71] Colombel JF, Mathieu D, Bouault JM, et al. Hyperbaric oxygenation in severe perineal Crohn's disease. *Dis Colon Rectum* 1995;38(6):609-14.
- [72] Nelson EW Jr, Bright DE, Villar LF. Closure of refractory perineal Crohn's lesion: integration of hyperbaric oxygen into case management. *Dig Dis Sci* 1990;35(12):1561-5.
- [73] Lavy A, Weisz G, Adir Y, Ramon Y, Melamed Y, Eidelman S. Hyperbaric oxygen for perianal Crohn's disease. *J Clin Gastroenterol* 1994;19(3):202-5.
- [74] Brady CE 3rd, Cooley BJ, Davis JC. Healing of severe perineal and cutaneous Crohn's disease with hyperbaric oxygen. *Gastroenterology* 1989;97(3):756-60.
- [75] Buchman AL, Fife C, Torres C, Smith L, Aristizabal J. Hyperbaric oxygen therapy for severe ulcerative colitis. *J Clin Gastroenterol* 2001;33(4):337-9.
- [76] Gurbuz AK, Elbuken E, Yazgan Y, Yildiz S. A different therapeutic approach in patients with severe ulcerative colitis: hyperbaric oxygen treatment. *South Med J* 2003;96(6):632-3.
- [77] Shiratsuch H, Basson MD. Differential regulation of monocyte/macrophage cytokine production by pressure. *Am J Surg* 2005;190(5):757-62.
- [78] Granowitz EV, Skulsky EJ, Benson RM, et al. Exposure to increased pressure or hyperbaric oxygen suppresses interferon-gamma secretion in whole blood cultures of health humans. *Undersea Hyperb Med* 2002;29(3):216-25.
- [79] Rossignol DA, Rossignol LW. Hyperbaric oxygen therapy may improve symptoms in autistic children. *Med Hypotheses* 2006;67(2):216-28.
- [80] Rossignol DA, Rossignol LW, James SJ, Melnyk S, Mumper E. The effects of hyperbaric oxygen therapy on oxidative stress, inflammation, and symptoms in children with autism: an open-label pilot study. *BMC Pediatr* 2007;7(1):36.
- [81] Dickinson DA, Forman HJ. Glutathione in defense and signaling: lessons from a small thiol. *Ann N Y Acad Sci* 2002;973:488-504.